Association of Lipidomics Signatures in Blood with Clinical Progression in Preclinical and Prodromal Alzheimer's Disease

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14 Abstract.

- Background: Lipidomics may provide insight into biochemical processes driving Alzheimer's disease (AD) pathogenesis
 and ensuing clinical trajectories.
- Objective: To identify a peripheral lipidomics signature associated with AD pathology and investigate its potential to predict
 clinical progression.
- ¹⁹ **Methods:** We used Bayesian elastic net regression to select plasma lipid classes associated with the CSF pTau/A β_{42} ratio ²⁰ as a biomarker of AD pathology in preclinical and prodromal AD cases from the ADNI cohort. Consensus clustering of the
- selected lipid classes was used to identify lipidomic endophenotypes and study their association with clinical progression.
- **Results:** In the *APOE4*-adjusted model, ether-glycerophospholipids, lyso-glycerophospholipids, free-fatty acids, cholesterol
- esters, and complex sphingolipids were found to be associated with the CSF pTau/A β_{42} ratio. We found an optimal number
- of five lipidomic endophenotypes in the prodromal and preclinical cases, respectively. In the prodromal cases, these clusters
- differed with respect to the risk of clinical progression as measured by clinical dementia rating score conversion.
- **Conclusion:** Lipid alterations can be captured at the earliest phases of AD. A lipidomic signature in blood may provide a
- dynamic overview of an individual's metabolic status and may support identifying different risks of clinical progression.

Keywords: Alzheimer's disease, heterogeneity, lipidomics, risk assessment

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29 INTRODUCTION

Current diagnostic research criteria for the 30 early detection of Alzheimer's disease (AD) are 31 based on disease-defining biomarkers of amyloi-32 dosis, tauopathy, and neurodegeneration [1]. These 33 biomarkers, however, are not precise enough to 34 predict individual clinical trajectories and risk of 35 clinical conversion [2]. More recently, multi-omics 36 approaches have been studied to account for the 37 heterogeneity of clinical courses in AD and iden-38 tify different clinic-pathological endophenotypes 39 as a potential basis for personalized medicine 40 [3, 4]. 41

As one important example, lipidomics provides 42 insight into metabolic endophenotypes that may mod-43 ify the effect of AD pathology on neurodegeneration 44 and clinical trajectories. Thus, lipids are involved 45 in many downstream processes of AD pathology, 46 such as membrane remodeling, modulation of trans-47 membrane proteins, including amyloid-B protein 48 precursor (ABPP) and its secretases, maintaining 49 blood-brain barrier function, myelination, cell sig-50 naling, and inflammation. In addition, they may even 51 influence upstream events such as oxidative stress 52 pathways and alterations of energy balance [5, 6]. 53 Recent genetic studies supported the role of lipids 54 in AD pathogenesis even beyond the apolipopro-55 tein E ε 4 allele (APOE4), which is considered the 56 major genetic risk factor for late-onset sporadic 57 AD (LOAD) [7]. Genome-wide association studies 58 (GWAS) have identified associations between disease 59 status and several genes involved in lipid homeosta-60 sis, such as CLU (clusterin), SORL1 (sortilin-related 61 receptor 1), ABCA7 (ATP-binding cassette, sub-62 family A, member 7), and *PLD3* (phospholipase-D3) 63 [7] in addition to the microglia related PLCG2 (phos-64 pholipase C-gamma) [8]. 65

Our study used targeted lipidomics data from 66 the Alzheimer's Disease Neuroimaging Initiative 67 (ADNI) cohort to identify lipid alterations in the 68 blood associated with AD pathology biomarker, 69 namely cerebrospinal fluid (CSF) pTau/AB42 ratio, 70 in people with preclinical or prodromal AD. In 71 a secondary exploratory analysis, we determined 72 lipidomic endophenotypes within prodromal and 73 preclinical cases, respectively, using a consensus 74 clustering approach. We investigated whether these 75 lipidomic endophenotypes contributed to predicting 76 subsequent clinical progression as determined by 77 dementia rating score (CDR) conversion in preclini-78 cal and prodromal AD cases.

MATERIALS AND METHODS

Cohort overview

This study used data provided by the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (http://adni.loni.usc.edu). ADNI is a large, multicenter, longitudinal study of older adults launched in 2003 by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, the Food and Drug Administration, private pharmaceutical companies, and non-profit organizations. The study was designed to acquire serial neuroimaging, clinical and neuropsychological assessments, and other biologic markers to monitor the progression of mild cognitive impairment (MCI) and early AD. A full description of the study protocols and analytical methods are provided at (http://www.adni-info.org/).

The final cohort consisted of 529 participants from the ADNI cohort having a baseline diagnosis of either cognitively normal or mild cognitive impairment along with complete CSF- biomarkers, lipidomics, and body mass index (BMI) data. BMI values were sorted into three categories as follows: BMI_low (average weight): 18.5-24.9 or (underweight): <18.5, BMI_medium (overweight): 25-29.9 and BMI_high (at least moderately obese): > 30. We further classified our participants into three diagnostic groups based on their CSF pTau/AB42 status, such that the cognitively normal (CN) group represents cognitively normal participants with CSF pTau/A β_{42} below the cut-off (0.025) [9]. Preclinical and prodromal groups had CSF pTau/A β_{42} above the optimized cut-off and an initial diagnosis of cognitively normal and MCI, respectively.

APOE genotyping

At the baseline visit, blood samples were obtained from the participants, shipped to the central biomarker analysis lab at the University of Pennsylvania, and processed using an *APOE* genotyping kit, as further described (http://adni.loni.usc.edu/wp-content/uploads/2010/09/ADNI_GeneralProcedures Manual.pdf). For subsequent analysis, we coded participants' *APOE* genotype according to the presence of ε 4 allele present as follows; 0: no ε 4 allele, 1:1 or 2 ε 4 alleles.

CSF biomarkers measurements

CSF amyloid- β (1-42) (CSF A β_{42}) and CSF Phospho-Tau (181P) (CSF pTau) were measured 125

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using the fully automated Roche Elecsys® immuno-126 assay platform at the UPenn/ADNI Biomarker Labo-127 ratory. CSF biomarkers $A\beta_{42}$ and pTau/A β_{42} were 128 binary classified based on the optimized cut-offs 129 977 pg/ml and 0.025, respectively. These cut-offs 130 were determined on the ADNI cohort then vali-131 dated against the visual reads of amyloid-B PET, as 132 explained in [9]. 133

134 Lipidomics data

Targeted Lipidomics analysis was carried out on 135 the plasma samples from ADNI participants using 136 ultra-high-performance liquid chromatography cou-137 pled with chromatographic separation to characterize 138 isomeric and isobaric lipid species. Mass spectrom-139 etry analysis was performed on an Agilent (6490 140 OOO) mass spectrometer in positive ion mode with 141 dynamic scheduled multiple reaction monitoring 142 (MRM). The analysis was conducted following the 143 lipidomics protocol developed by Kevin Huynh and 144 Peter Meikle in Baker Heart and Diabetes Institute, 145 Metabolomics laboratory. A detailed description of 146 their lipidomics platform was provided in the method-147 ology file (ADNI_ADMCLIPIDOMICSMEIKLEL 148 ABLONG_METHODS_20210121.pdf) and respec-149 tive articles [10,11]. 150

After applying the standard normalization and 151 batch correction procedures, measurements from 692 152 lipid species were provided in the file (ADMCLIPI 153 DOMICSMEIKLELABLONG.csv). All the lipid 154 measurements were log10 and z-transformed before 155 any analysis. Lipid species (692) were then merged 156 into one hundred and seven (107) composite scores 157 defined through a hierarchical clustering approach 158 that was applied within each of the lipid subclasses 159 /classes. 160

161 Statistical analysis

Selection of salient lipids associated with biomarkers of AD pathology

We used Bayesian elastic net regularized logistic 164 regression to select lipid composite scores associated 165 with the CSF pTau/A β_{42} ratio as a biomarker of AD 166 pathology. Regularized logistic regression methods 167 were developed to carry out simultaneous parame-168 ter estimation and variable selection [12, 13]. Elastic 169 net offers an optimum regularization and variable 170 selection, particularly in high dimensional data set-171 tings, such as the current lipidomics data, where 172 features are often highly collinear, and their num-173 ber exceeds the sample size [13, 14]. As one of 174

the regularization approaches, the elastic net provides a reasonable compromise between both ridge (L2) and lasso (L1) penalties [13, 14]. It performs an effective feature selection via the lasso penalty while better handling correlated features via the ridge penalty [14, 15]. Adopting a Bayesian approach possesses several advantages over classic elastic net regularized regression [12, 16]. First, Bayesian methods provide a straightforward statistical inference for the estimated coefficients through the posterior distributions and credibility intervals [12, 16]. Second, it allows for simultaneous estimation of both penalty parameters (L2 & L1) and model parameters [12, 16]. This is particularly important in controlling the double shrinkage problem (too small, estimated coefficients) due to sequential estimation of penalty parameters through cross-validation procedure in the classic method. Additionally, Bayesian approaches have shown better variable selection in real data examples and simulation studies [12].

Before conducting the analysis, lipid composite 195 scores were transformed into W-scores using regres-196 sion models estimated on the control group. W-scores 197 are analogous to Z-scores yet adjusted for particular 198 covariates, namely age and sex [17]. An initial filter-199 ing step was carried out to include only the top 60% 200 of lipid composite scores correlated with the CSF 201 pTau/AB42 status in the regularized logistic regres-202 sion models. Then, a Bayesian logistic regression 203 model with elastic net regularization was fitted in 204 the RStan interface. We adapted the scripts provided 205 by Sara van Erp on GitHub (https://github.com/sara-206 vanerp/bayesreg), implementing elastic net priors in 207 Bayesian regularized regression models using Stan 208 language [16]. A training dataset (80% of the whole 209 cohort) was used for estimation of model parame-210 ters through Markov Chain Monte Carlo (MCMC) 211 sampling (No-U-Turn Sampler (NUTS) algorithm). 212 The resulting estimates were then used to predict 213 the outcome in the test dataset (20 % of the whole 214 cohort). Lipid composite scores were selected based 215 on the credible interval criterion, where a variable is 216 excluded if the credibility interval covers 0. A credi-217 bility interval level of 50% was used as recommended 218 in [12]. Salient lipid composite scores were deter-219 mined based on being selected in more than 50% 220 of the cross-validation 100 iterations. Three different 221 models were calculated: 1) Reference model, using 222 the demographic criteria (Age and Sex); 2) Lipid 223 model, using lipid composite W-scores, and 3) Lipid 224 model + APOE4, where participants' APOE4 status 225 was added as a covariate to the Lipid model.

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Prediction of clinical progression

Lipidomic endophenotypes based on consensus 227 clustering. We applied a hierarchical clustering on 228 those lipid composite scores that had been found 229 associated with the CSF pTau/AB42 ratio in the 230 previous regularized regression analysis. The clus-231 tering was performed separately in the preclinical 232 and prodromal subgroups, respectively. We employed 233 a consensus clustering approach using data sub-234 sampling [18, 19], repeated 5,000 times to ensure 235 the stability and robustness of clustering results. 236 During each repetition, 80% of the data samples 237 (participants) were randomly selected for agglomer-238 ative hierarchical clustering using Ward's criterion 239 to minimize the total within-cluster variance. A con-240 sensus matrix/cluster-based similarity matrix was 241 then constructed. Each element in the matrix is a 242 number between 0 and 1 inclusive, representing 243 the proportion of times that two samples (partic-244 ipants) were clustered together out of the times 245 that the same samples were chosen in the bootstrap 246 sub-sampling process. Then final cluster assignment 247 was defined through the consensus function, cluster-248 based similarity partitioning algorithm (CSPA), first 249 introduced by Strehl and Ghosh and implemented 250 in diceR library [18]. CSPA is an efficient con-251 sensus function that re-clusters the data samples 252 through applying hierarchical clustering on the 253 constructed consensus matrix [18, 19]. Hence the 254 cluster labels are inferred at the hierarchy level 255 of the optimal number of clusters (k) previously 256 defined. 257

The optimal number of clusters was defined based 258 on a composite score combining the proportion of 259 ambiguous clustering (PAC) score and Dunn's index 260 estimated within the consensus clustering. PAC is 261 a robust estimate of cluster stability, mainly when 262 data samples are not independent [20], an intrinsic 263 feature of omics data. PAC score is the fraction of 264 sample pairs with consensus index values falling in 265 the intermediate interval, i.e., PAC window. In a per-266 fect clustering, the consensus matrix would consist 267 of zeros or ones, and therefore the PAC score would 268 be zero [20]. Thus, the lower the PAC score, the more 269 stable and near perfect the clusters. We used a PAC 270 window of (0.1, 0.9) in our analysis. 271

Conversely, Dunn's index estimates clustering
internal validity considering compactness and separation measures [21]. The larger the Dunn's index, the
better the inter-cluster separability and intra-cluster
compactness. The composite score was computed
as PAC score divided by Dunn's index value;

accordingly, the lower the composite score, the better the clustering.

Lipidomic endophenotypes and risk of CDR conversion. We assessed the potential of the defined lipidomic endophenotypes to predict Clinical Dementia Rating score (CDR) conversion from a value of 0 to 0.5 or 0.5 to 1 or higher in the preclinical and prodromal sub-cohorts, respectively. Using Bayesian survival analysis, we estimated the risk of conversion over a follow-up period of six years (average follow-up = 4.15 + 1.72) while accounting for censoring. We further explored the effect of several covariates, namely age, sex, BMI, APOE4, and years of education, on the estimated risk of conversion. Finally, Bayesian multivariate analysis (MANOVA) was conducted to reveal which lipid composite scores distinguished clusters at low versus high risk of clinical progression.

The whole analysis workflow is summarized in Fig. 1. All analyses were performed in R (version 3.6.3) using the following packages: RStan (version 2.21.2), RStanArm, brms, bayestestR, BayesFactor, pROC, diceR.

RESULTS

Demographic characteristics

A summary of the demographic characteristics of our final cohort is provided in (Table 1). The diagnostic groups did not differ in age, sex, or education years. The distribution of BMI categories differed between groups; the preclinical group had the highest proportion of BMI-low category. As expected, the *APOE* ε 4 allele was more prevalent in preclinical and prodromal groups (\geq 60%) compared with the normal control group (pTau/A β_{42} -ve) (18%). AD CSF biomarker levels (pTau and pTau/A β_{42}) were higher in prodromal participants than in the preclinical group.

Selection of salient lipids associated with biomarkers of AD pathology

Bayesian elastic net regularized logistic regression models performance

Using only age and sex as predictors, the performance of the Reference model was not better than random prediction. The Lipid model improved the prediction accuracy. The cross-validated area under the receiver operating curves (CV-AUC), CV-Accuracy, CV-Sensitivity, and CV-Specificity at the optimum threshold were 0.65, 0.66, 0.68, and 0.61,

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Fig. 1. Overview of the data analysis workflow. This figure summarizes the analysis workflow adopted by this study as described in the Materials and Methods section. Panel A displays the preparation of the final cohort based on the defined inclusion criteria then the classification of the final diagnostic groups based on the CSF pTau/A β_{42} ratio. The statistical analysis is demonstrated in panels B and C. Panel B illustrates the selection of salient lipids associated with biomarkers of AD pathology through Bayesian elastic net regularized logistic regression models. Panel C explains the steps to predict clinical progression in the diagnostic groups, namely prodromal and preclinical. First, we defined clusters of participants having similar lipid profiles within each diagnostic group. Then we explored the defined clusters for the risk of conversion to MCI or dementia.

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	Overview of cohort demographics								
	CN	Preclinical	Prodromal	Whole cohort					
N	182	73	274	529					
Mean age (sd) ^a	73.2 (5.9)	75.9 (5.2)	73.3 (7.0)	73.6 (6.5)					
Sex – Females ^b N (percent %)	88 (48 %)	41 (56 %)	109 (40 %)	238 (45 %)					
APOE4 carriers ^{b***} N (percent %)	32 (18 %)	43 (59 %)	195 (71 %)	270 (51 %)					
BMI ^{b***} N (percent %)									
Low	50 (27%)	38 (52%)	113 (41%)	201 (38%)					
Medium	85 (47%)	21 (29%)	126 (46%)	232 (44%)					
High	47 (26 %)	14 (19 %)	35 (13%)	96 (18%)					
Mean Education y (sd) ^a	16.3 (2.7)	16.0 (2.8)	15.9 (2.9)	16.1 (2.8)					
CSF biomarkers									
Mean A β_{42} (sd) ^{a***}	1727.0 (524.0)	634.0 (185.0)	630.0 (167.0)	1007.8 (620.4)					
Mean pTau (sd) ^{a***}	20.1 (6.6)	28.8 (10.4)#	35.4 (14.1)#	29.2 (13.4)					
Mean pTau/A β_{42} ratio (sd) ^{a***}	0.012 (0.003)	0.049 (0.025)#	0.059 (0.028)#	0.042 (0.03)					

Summary of the demographic characteristics of our cohort split into the final three diagnostic groups cognitively normal elderly (*CN*), preclinical and prodromal. Characteristics are described as Number (N) and the corresponding percentage (percent %) or Mean value and standard deviation (sd) as convenient. Group differences were tested using Bayesian ANOVA (a) and Bayesian test of association (b). Results were interpreted in terms of Bayes Factor (BF) in favor of presence of group differences in the tested variables, where BF of (3–20) represented moderate evidence (*), BF of (20–150) represented strong evidence (**) while BF of (>150) represented very strong evidence (***). Differences in levels of CSF biomarkers levels between Preclinical and Prodromal are marked by ([#]).

respectively. However, the best performance was achieved by the Lipid+*APOE4* model; the estimated CV-AUC, CV-Accuracy, CV-Sensitivity, and CV-Specificity increased to 0.76, 0.71, 0.69, and 0.77, respectively. Supplementary Table 1 provides an overview of all tested models.

332 Identification of salient lipids

The Lipid + *APOE4* model selected a set of twentyeight lipid composite scores in at least 50% of cross-validation repetitions (Supplementary Table 2). A features' relative importance and stability were determined by the median posterior β -coefficients and frequency of selection across the cross-validations. According to these criteria, lyso-glycero-phospholipids (LPL), alkenyl-glycerophospholipids (plasmalogens), free fatty acids (FFA), cholesterol esters and sphingolipids (complex ceramides) lipid classes/subclasses ranked on top of the list. Both lyso-phosphotipide (LPC_7: poly-unsaturated fatty

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acid (PUFA)) and lyso-alkyl-phosphatidylcholine 345 (LPC_O_2: long-chain fatty acid (FA)) were posi-346 tively associated with the CSF pTau/ AB42 ratio. 347 Similarly, phosphatidylcholine (PC_5: arachidonic 348 acid (AA)) harboring arachidonic acid showed a pos-349 itive association. Conversely, plasmalogens such as 350 alkenyl- phosphatidylcholine (PC_P_5: docosahex-351 aenoic acid (DHA), Eicosapentaenoic acid (EPA) & 352 PC_P_2: saturated and mono-unsaturated FA) and 353 alkenyl- phosphatidylethanolamine (PE_P_5: AA, 354 DHA) showed negative associations. 355

Except for AA (FA_3), free fatty acids (FA_1: sat-356 urated, mono-unsaturated, PUFA) were negatively associated with the AD biomarkers. Cholesterol 358 esters (Chols_ester_3: PUFA & Chols_ester_2) and 359 long-chain acyl-carnitines (AC_4: PUFA) were 360 positively associated with AD biomarkers, while di-acylglycerol (DG_3: EPA, DHA) and alkyl-di-362 acylglycerol (TG_O_3) showed negative relation. 363

Complex ceramides including hexosyl-ceramides 364 (hexCER_6 & hexCER_7), gangliosides (GM1), and 365 sulfatides were found to be positively associated with 366 AD biomarkers yet di-hydro-ceramides (dhCER_1), 367 gangliosides (GM3_3: very long FA), and sphin-368 gomyelin (SM_3: very long FA) were negatively 369 associated. Figure 2 displays the median posterior B-370 coefficients and their credibility intervals across the 371 cross-validations, as estimated by the Lipid + APOE4 372 model. Lipid species, constituting each of the salient 373 lipid composite scores, are listed in Supplementary 374 Table 3. 375

Prediction of clinical progression 376

Lipidomic endophenotypes based on consensus 377 clustering 378

We conducted consensus clustering to identify 379 lipidomic endophenotypes based on the set of lipid 380 composite scores selected by the Lipid+APOE4 381 model. 382

In the prodromal sub-cohort, we determined the 383 optimum number of clusters to be (k=5), as demon-384 strated in Supplementary Figure 1. Of the prodromal 385 participants, 28% fell into the cluster (I), 23% in the 386 cluster (IV), 20% each in the clusters (II) and (V), 387 and 9% in the cluster (III). Apart from the BMI cate-388 gories distribution, there was no conclusive evidence 389 for differences in age, sex, years of education, APOE4 390 status, or the CSF levels of AD biomarkers between 391 the defined clusters (Supplementary Table 4). 392

Following the same approach, we determined (k =393 5) the optimal number of clusters for the preclinical 394

sub-cohort, as shown in Supplementary Figure 2. Of these participants, 28% fell into the cluster (I), while the rest were equally distributed over the remaining clusters. Details on the distribution of demographic characteristics, APOE4 genotype, and BMI categories can be found in Supplementary Table 5.

Lipidomic endophenotypes and risk of CDR conversion

We evaluated the risk of CDR conversion among prodromal sub-cohort clusters with and without adjusting for the effect of covariates as demonstrated in Supplementary Table 6. Cluster (IV) was chosen as the reference group since it exhibited a lower risk of CDR conversion. Moreover, cluster (IV) enclosed a relatively large proportion of participants. As shown in Fig. 3, the clusters (II) (HR = 1.97 (1.26 - 3.10)) and (V) (HR = 1.99 (1.30-3.00)) had an increased risk of conversion in the APOE4 adjusted model. To investigate whether these effects differed between sexes, we repeated the Bayesian survival models (APOE4 adjusted) in the male and female data subsets, respectively (Table 2). In men, the lipid profiles of clusters (II and V) showed an increased risk of conversion, whereas cluster (III) showed a decreased risk of conversion relative to the reference cluster (IV). In women, only cluster (II) had an increased risk of conversion.

Finally, we conducted Bayesian multivariate analysis to identify differences in lipid composite scores between the reference cluster (IV) and the remaining clusters (Supplementary Table 7). Figure 4 shows the specific lipid profile for each cluster of the prodromal sub-cohort.

In the preclinical sub-cohort, there was no evidence of a difference in risk of CDR conversion between the five clusters. Essentially identical results were obtained whether we adjusted or not for covariates.

DISCUSSION

We explored different lipid classes in preclinical and prodromal AD cases to analyze the relationship between lipid metabolism markers and biomarkers of amyloid and tau pathology, as well as clinical progression.

Our first goal was to determine associations between peripheral lipid alterations and pathology markers of AD in the CSF. Ether glycerophospholipids, particularly plasmalogens, showed lower levels in preclinical and prodromal AD participants 399

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Fig. 2. Salient lipids associated with CSF pTau/Aβ₄₂ ratio. We used Bayesian elastic net logistic regression (Lipid+*APOE4* model) to select salient lipid composite scores associated with CSF pTau/Aβ₄₂ ratio. Estimated posterior β-coefficients are represented as points with their respective 50% and 90% credibility intervals as thick and thin error bars, respectively. The points' color codes for their corresponding lipid class. LPC_O_2: Lyso-alkyl-phosphatidylcholine (long/ very long FA), Choles_ester_3: Cholesteryl ester (PUFA), hexCER: Hexosyl-ceramide, FA_3: Free fatty acid (AA), PC_5: Phosphatidylcholine (AA), LPC_7: Lysophosphatidylcholine (PUFA), AC_4: Acylcarnitine (PUFA), GM1: GM1 gangliosides, Choles_ester_2: Cholesteryl ester, SULF_1: Sulfatides, LPE_1: Lyso-phosphatidylethanolamine (saturated FA), PL_1: Phosphatidylinositol (PUFA), LPL_3: Lyso-phosphatidylinositol (AA), GM3_3: GM3 gangliosides (very long FA), dhCER: Dihydroceramide, LPC_P_2: Lyso-alkenyl-phosphatidylcholine (long FA), SM_3: Sphingomyelin (very long saturated FA), PL_2: Phosphatidylinositol (saturated, monounsaturated FA), LPC_5: Lysophosphatidylcholine (long, very long FA), LPC_2: Lysophosphatidylcholine (odd numbered FA), TG_O_3: Alkyl-diacylglycerol, DG_3: diacylglycerol (EPA & DHA), PC_P_2: Alkenyl-phosphatidylcholine (saturated and mono-unsaturated FA), PE_P_5: Alkenyl-phosphatidylethanolamine (AA, DHA), PC_P_5: Alkenyl-phosphatidylcholine (DHA & EPA) and FA_1: Free fatty acid.

compared with controls. Conversely, we found ara-443 chidonic acid-containing phosphatidylcholine, 444 PUFA (omega-3) lyso-phosphatidylcholine and lyso-445 alkyl-phosphatidylcholine with predominant satu-446 rated/mono-unsaturated long-chain fatty acid to be 447 increased. Low levels of plasmalogens have been 448 frequently linked to AD pathology [22], whether 449 measured in brain tissue [23-25], CSF [25], or 450 plasma blood samples [26]. Grey matter plasmalo-451 gens (DHA and AA at sn-2) depletion was found 452 associated with disease progression and severity in 453

AD patients [27–30]. A recent study by Lim et al. proposed that ether-lipids dysregulation may partly mediate the effect of two major AD risk factors, namely, age and *APOE4* [31].

Toledo et al. showed that higher baseline levels of long-chain and PUFA-containing alkyl phosphatidylcholines (PC ae 42:4, PC ae 44:4) correlated with abnormal levels of CSF $A\beta_{42}$ in preclinical and prodromal AD participants of the ADNI cohort and predicted conversion from MCI to AD dementia [32]. In the current study, we observed high levels

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Fig. 3. Lipid endophenotypes predict clinical progression to dementia. We conducted a Bayesian survival analysis to estimate the risk of clinical progression to dementia among the pre-defined clusters of the prodromal sub-cohort. Clinical progression in the prodromal sub-cohort is defined as the conversion of clinical dementia rating score (CDR) from a value of 0.5 to 1. Clusters (II and V) are found to have $\simeq 2$ folds higher risk of progression to dementia compared to the reference cluster (IV).

Model	Cluster + APOE4			Male subset			Female subset		
	Median (MAD)	Hazard ratio	HDI	Median (MAD)	Hazard ratio	HDI	Median (MAD)	Hazard ratio	HDI
Intercept: IV	-9.03 (1.80)			-8.46 (2.20)				-9.24 (2.72)	
I	0.02 (0.26)	1.02	0.68-1.52	-0.16 (0.32)	0.85	0.54-1.51	0.28 (0.42)	1.33	0.68-2.72
II	0.68 (0.28)	1.97	1.26-3.10	0.56 (0.35)	1.75	1.04-3.16	0.84 (0.43)	2.32	1.15-4.57
III	-0.41 (0.42)	0.66	0.36-1.22	-1.08(0.58)	0.34	0.13-0.89	0.09 (0.54)	1.10	0.48-2.56
V	0.69 (0.26)	1.99	1.30-3.00	0.85 (0.34)	2.35	1.38-4.06	0.55 (0.43)	1.74	0.89-3.53
APOE4	0.39 (0.21)	1.48	1.07-2.05	0.40 (0.27)	1.50	1.00 - 2.25	0.27 (0.33)	1.31	0.76-2.23

Table 2 Risk of clinical progression among prodromal lipidomic endophenotypes

Bayesian survival analysis was conducted to estimate the relative risk of progression to dementia among prodromal lipidomic endophenotypes while adjusting for *APOE4* adjusted model was selected based on the sensitivity analysis provided in Supplementary Table 6, which investigated the relative risk of several covariates. We further replicated the same model on male and female subsets separately to explore sex-specific effect of lipidomic endophenotypes on clinical progression. Throughout the analysis, we set cluster (IV) as our reference group. Results were interpreted in terms of high-density intervals (HDI) of posterior distributions, where hazard ratios with HDI not covering (1) were considered relevant and reported in red.

of arachidonic acid-containing phosphatidylcholine, and long-chain alkyl lyso-phosphatidylcholines (LPC-O), were associated with the CSF pTau/Aβ₄₂ ratio. Results from both studies suggest an early role of arachidonated phosphatidylcholines, particularly long-chain alkyl isomers and their lyso derivatives, in AD pathogenesis, even in cognitively normal individuals with pathological levels of CSF AD biomarkers. These phosphatidylcholine species are known precursors of potent inflammatory mediators, including platelet-activating factor (PAF) and arachidonic acid. Additionally, they are highly abundant in platelets and immune cells [33, 34]. This points to a potential regulatory role in inflammation processes and would represent a possible link between inflammation and AD [32].

Complex ceramides, including glycosylated ceramides, GM1 gangliosides, and their precursors 481

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Fig. 4. Heterogeneity of lipidomic endophenotypes among the prodromal sub-cohort. The specific lipid profile of each cluster is demonstrated on a heatmap in terms of average w-scores. On the color scale, red represents scores higher than expected in the age and sex-matched control group, and blue color represents lower scores. Bayesian multivariate analysis was conducted to identify lipid composite scores distinguishing clusters at higher risk of clinical progression from the reference group. Cluster (IV) was set as the reference group and marked by (Ref.). Clusters (II and V) were defined as groups at higher risk of progression and marked by (#). Asterisk (*) points to lipid scores that showed evidence of group differences. PC_5: Phosphatidylcholine (AA), PC_P_2: Alkenyl-phosphatidylcholine (saturated and mono-unsaturated FA), PC-P_5: Alkenyl-phosphatidylcholine (DHA & EPA), PE_P_5: Alkenyl-phosphatidylethanolamine (AA, DHA), PI_1: Phosphatidylinositol (PUFA), PI_2: Phosphatidylinositol (saturated, monounsaturated FA), LPC_2: Lysophosphatidylcholine (odd numbered FA), LPC_5: Lysophosphatidylcholine (long, very long FA), LPC_7: Lysophosphatidylcholine (PUFA), LPC_0_2: Lyso-alkylphosphatidylcholine (long/very long FA), LPC_P_2: Lyso-alkenyl-phosphatidylcholine (long FA), LPE_1: Lyso-phosphatidylethanolamine (saturated FA), LPI_3: Lyso-phosphatidylinositol (AA), dhCER: Dihydroceramide, hexCER: Hexosyl-ceramide, GM3_3: GM3 gangliosides (very long FA), GM1: GM1 gangliosides, SM_3: Sphingomyelin (very long saturated FA), SULF_1: Sulfatides, Choles_ester_2: Cholesteryl ester, Choles_ester_3: Cholesteryl ester (PUFA), DG_3: diacylglycerol (EPA & DHA), TG_0_3: Alkyl-diacylglycerol, FA_1: Free fatty acid, FA_3: Free fatty acid (AA) and AC_4: Acylcarnitine (PUFA).

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hexosyl-ceramides and sulfatides, showed higher lev-483 els in prodromal and preclinical AD participants, in 484 contrast to di-hydro-ceramides, sphingomyelins, and 485 GM3 gangliosides, which were decreased. Several 486 studies suggested a shift in sphingolipids metabolism 487 towards ceramides accumulation [35, 36] and deple-488 tion of sphingomyelins, particularly those with 489 long-chain FA (C22, C24) [37, 38] and sulfated 490 sphingolipids [35] early in the course of AD [39]. 491 Ceramides, a key bioactive molecule in sphingolipids 492 metabolism, were suggested to contribute to the 493 increased susceptibility of neurons and oligodendro-494 cytes to apoptotic cell death [40]. This hypothesis 495

W – scores

was further supported by the elevated activity of enzymes involved in ceramides synthesis, namely sphingomyelinases and ceramidases, in brain tissue of AD cases [38]. Consistent with these findings, gene expression of sphingomyelinases and serine palmitoyl transferase enzymes was found to be upregulated in AD patients' brain tissue [36, 39].

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AC_4

The second goal of our study was to identify distinct lipidomic endophenotypes and assess their association with clinical progression. Lipidomics endophenotyping offers a global mapping of the alterations in biochemical pathways [41]. These alterations may partly reflect underlying AD pathology.

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Additionally, these endophenotypes can capture com-509 plementary information related to an individual's 510 specific comorbidities and/or genomic characteris-511 tics that could partly explain the diversity observed 512 in clinical trajectories within AD populations [3]. In 513 the prodromal sub-cohort, the lipid profiles of clus-514 ters (II and V) were associated with a higher risk of 515 clinical progression. In both clusters, we observed 516 lower levels of PUFA (mainly AA) containing plas-517 malogens and phosphatidylcholines associated with 518 a compensatory increase of plasmalogens, mainly 519 alkenyl phosphatidylcholines, containing saturated 520 and mono-unsaturated FAs. Higher levels of choles-521 terol esters, complex ceramides together with the 522 depletion of long-chain sphingomyelins, and di-523 hydro-ceramides were also noted in clusters (II and 524 V) participants. Cluster (III) lipidomic profile was 525 associated with a lower risk of progression (CDR con-526 version) yet only in men. Cluster (III) constituted a 527 group of prodromal participants with a higher preva-528 lence of low BMI and a slightly higher proportion of 529 APOE4 carriers compared with the reference cluster 530 (IV). 531

Previous studies used logistic regression or 532 machine learning algorithms to investigate the asso-533 ciation of lipids with dementia risk in cognitively 534 normal individuals [42-44] and people with MCI 535 [32, 45]. Several studies have found higher levels of 536 sphingomyelin, phosphatidylcholines, and lysophos-537 phatidylcholine associated with conversion from 538 MCI to AD/dementia [32, 46, 47]. Conversely, Map-539 stone et al. [43] and Ma et al. [45] showed that 540 lower baseline levels of phosphatidylcholines and 541 lysophosphatidylcholine were significantly associ-542 ated with accelerated cognitive decline [45] and risk 543 of conversion to MCI/AD compared to cognitively 544 stable participants [43]. 545

In a different approach, Wood et al. [48] addressed 546 heterogeneity in lipid alterations patterns within 547 groups of MCI and AD cases. They defined sub-548 groups within each diagnostic group according to 549 their Mini-Mental State Examination score (low ver-550 sus high). Based on the literature, they focused on two 551 lipid classes, ethanolamine plasmalogens and diacyl-552 glycerols. MCI and AD cases had elevated levels 553 of diacylglycerols and plasmalogens depletion com-554 pared with controls [48]. Low and high Mini-Mental 555 State Examination MCI cases, however, showed no 556 differences in both lipid classes [48]. In contrast to 557 such a hypothesis-driven approach, here we explored 558 the diversity of lipidomic endophenotypes within 559 prodromal cases using an unsupervised clustering 560

approach. Thus, our findings serve to generate rather than confirm hypotheses on the association of lipid profiles with the risk of conversion.

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Recent evidence suggested that sex has an effect on the association of lipids with AD pathology and rates of cognitive decline [31, 49, 50]. In our study, cluster (III) showed a decreased risk of conversion in men but not in women. This cluster had high levels of longchain fatty acids lysophosphatidylcholine (both acyl and ether) and plasmalogens together with low levels of acylcarnitines. Sex-specific remodeling of lipid metabolism was suggested before, where high levels of sphingomyelins and phosphatidylcholines were reported in women [49, 50]. Conversely, lysophosphatidylcholine and ceramides were found at higher levels in men [49]. Thus, phospholipases may have higher activity in men and sphingomyelin synthetase may have a higher activity in women [49]. Consequently, we adjusted lipid scores for age and sex based on the control group in an attempt to control for the complex interaction of lipids with sex during different stages of AD. Although we started with a substantial number of cases, the sample size within preclinical and prodromal sub-cohorts and their respective lipid endophenotypes clusters was small, so that it was not feasible to conduct the full analysis in a sex-stratified fashion, as recommended in [49, 50].

Lack of consistency across metabolomics studies' results always was and still is a major limitation that hinders including lipid markers into diagnostic biomarker panels of AD [50, 51]. This heterogeneity is related to many factors, among them variability in data processing procedures and analytical platforms [51], as well as studies' design, sample size, distribution of relevant risk factors, and used statistical approaches [50]. Another factor probably is the lack of strong effects which contributes to inconsistent findings across studies. In our Bayesian regression models, we observed overall small contributions from individual lipid composite scores to the association with AD pathology CSF biomarkers as indicated by poor model performance as well as small posterior coefficients with large credibility intervals. In addition, metabolomics data are inherently highly collinear. This could contribute to high variance observed within the models and difficulty assessing variables' relative importance [52]. Taken together, a wide range of variance is observed in metabolomics data that limits their integration in the first line of diagnostic workflow and renders them likely more useful in adding to the accuracy of other prognostic markers [48].

Several limitations need to be acknowledged in 613 this study. Instead of using raw lipid scores, we used 614 composite scores based on hierarchical clustering 615 applied within each lipid class. Such an approach 616 could have masked the effects of some individual 617 lipid species. Our objective was to reduce data dimen-618 sionality and overcome the drawback of variables' 619 multicollinearity, particularly on regression coeffi-620 cients estimation and model stability. Concurrently 621 we wanted to maintain the representation of all inves-622 tigated lipid subclasses/classes and identify subsets 623 of functionally similar lipid species. Finally, given 624 the heterogeneity of lipidomics data, particularly in 625 early AD individuals, even larger cohorts are needed 626 to identify endophenotypes robustly. In future anal-627 ysis, we would like to tune and then validate our 628 approach on a larger sample derived from multiple 629 cohorts and particularly enriched with participants in 630 the preclinical stage of AD. 631

632 CONCLUSION

Through our study, we have shown that alter-633 ations in lipids, particularly those harboring poly-634 unsaturated fatty acids and ether bonds, can be 635 captured at the earliest stages of AD. Lipidomics pro-636 files provide an overview of an individual's metabolic 637 status whilst incorporating the balance within and 638 between interacting biochemical pathways. Hence, 639 identifying distinct lipidomic endophenotypes could 640 contribute to AD risk and clinical trajectories. Refin-641 ing and validating this approach could open a new 642 avenue to adjuvant interventions modulating lipid 643 metabolic pathways and allow for targeting subjects 644 with the largest expected benefit. 645

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SUPPLEMENTARY MATERIAL

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REFERENCES

- [1] Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, Iwatsubo T, Jack CR, Kaye J, Montine TJ, Park DC, Reiman EM, Rowe CC, Siemers E, Stern Y, Yaffe K, Carrillo MC, Thies B, Morrison-Bogorad M, Wagster M V., Phelps CH (2011) Toward defining the preclinical stages of Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* **7**, 280-292.
- [2] Dumurgier J, Hanseeuw BJ, Hatling FB, Judge KA, Schultz AP, Chhatwal JP, Blacker D, Sperling RA, Johnson KA, Hyman BT, Gómez-Isla T (2017) Alzheimer's disease biomarkers and future decline in cognitive normal older adults. *J Alzheimers Dis* 60, 1451-1459.
- [3] Badhwar AP, McFall GP, Sapkota S, Black SE, Chertkow H, Duchesne S, Masellis M, Li L, Dixon RA, Bellec P (2020) A multiomics approach to heterogeneity in Alzheimer's disease: Focused review and roadmap. *Brain* 143, 1315-1331.

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- [4] Hampel H, O'Bryant SE, Molinuevo JL, Zetterberg H, Masters CL, Lista S, Kiddle SJ, Batrla R, Blennow K (2018)
 Blood-based biomarkers for Alzheimer disease: Mapping the road to the clinic. *Nat Rev Neurol* 14, 639-652.
- [5] Chew H, Solomon VA, Fonteh AN (2020) Involvement of lipids in Alzheimer's disease pathology and potential therapies. *Front Physiol* 11, 598.
- [6] Wong MW, Braidy N, Poljak A, Pickford R, Thambisetty M, Sachdev PS (2017) Dysregulation of lipids in Alzheimer's disease and their role as potential biomarkers. *Alzheimers Dement* 13, 810-827.
- [7] Giri M, Zhang M, Lü Y (2016) Genes associated with Alzheimer's disease: An overview and current status. *Clin Interv Aging* 11, 665-681.
- [8] Magno L, Lessard CB, Martins M, Lang V, Cruz P, Asi Y, Katan M, Bilsland J, Lashley T, Chakrabarty P, Golde TE, Whiting PJ (2019) Alzheimer's disease phospholipase C-gamma-2 (PLCG2) protective variant is a functional hypermorph. *Alzheimers Res Ther* 11, 16.
- [9] Hansson O, Seibyl J, Stomrud E, Zetterberg H, Trojanowski JQ, Bittner T, Lifke V, Corradini V, Eichenlaub U, Batrla R, Buck K, Zink K, Rabe C, Blennow K, Shaw LM (2018) CSF biomarkers of Alzheimer's disease concord with amyloid-β PET and predict clinical progression: A study of fully automated immunoassays in BioFINDER and ADNI cohorts. *Alzheimers Dement* 14, 1470-1481.
- [10] Huynh K, Barlow CK, Jayawardana KS, Weir JM, Mellett NA, Cinel M, Magliano DJ, Shaw JE, Drew BG, Meikle PJ (2019) High-throughput plasma lipidomics: Detailed mapping of the associations with cardiometabolic risk factors. *Cell Chem Biol* 26, 71-84.
- [11] Huynh K, Lim WLF, Giles C, Jayawardana KS, Salim A, Mellett NA, Smith AAT, Olshansky G, Drew BG, Chatterjee P, Martins I, Laws SM, Bush AI, Rowe CC, Villemagne VL, Ames D, Masters CL, Arnold M, Nho K, Saykin AJ, Baillie R, Han X, Kaddurah-Daouk R, Martins RN, Meikle PJ (2020) Concordant peripheral lipidome signatures in two large clinical studies of Alzheimer's disease. *Nat Commun* 11, 5698.
- [12] Li Q, Lin N (2010) The Bayesian elastic net. *Bayesian Anal* **5**, 151-170.
- [13] Zou H, Hastie T (2005) Regularization and variable selection via the elastic net. J R Stat Soc B Stat Methodol 67, 301-320.
- [14] Hastie T, Tibshirani R, Wainwright M (2015) Statistical learning with sparsity. In *Statistical Learning with Sparsity: The Lasso and Generalizations*, Chapman and Hall/CRC, pp. 55-93.
- [15] Kuhn M, Johnson K (2013) Applied Predictive Modeling, Springer New York, New York.
- [16] van Erp S, Oberski DL, Mulder J (2019) Shrinkage priors for Bayesian penalized regression. J Math Psychol 89, 31-50.
- [17] Dyrba M, Mohammadi R, Grothe MJ, Kirste T, Teipel SJ
 (2020) Gaussian graphical models reveal inter-modal and
 inter-regional conditional dependencies of brain alterations
 in Alzheimer's disease. *Front Aging Neurosci* 12, 99.
- [18] Strehl A, Ghosh J (2002) Cluster ensembles A knowledge
 reuse framework for combining multiple partitions. *J Mach Learn Res* 3, 583-617.
- [19] Hennig C, Meila M, Murtagh F, Rocci R (2015) *Handbook of Cluster Analysis*, Chapman and Hall/CRC.
- [20] Şenbabaoğlu Y, Michailidis G, Li JZ (2014) Critical limita tions of consensus clustering in class discovery. *Sci Rep* 4,
 6207.

- [21] Dunn JC (1974) Well-separated clusters and optimal fuzzy partitions. *J Cybern* **4**, 95-104.
- [22] Su XQ, Wang J, Sinclair AJ (2019) Plasmalogens and Alzheimer's disease: A review. *Lipids Health Dis* 18, 100.
- [23] Ginsberg L, Rafique S, Xuereb JH, Rapoport SI, Gershfeld NL (1995) Disease and anatomic specificity of ethanolamine plasmalogen deficiency in Alzheimer's disease brain. *Brain Res* 698, 223-226.
- [24] Grimm MOW, Kuchenbecker J, Rothhaar TL, Grösgen S, Hundsdörfer B, Burg VK, Friess P, Müller U, Grimm HS, Riemenschneider M, Hartmann T (2011) Plasmalogen synthesis is regulated via alkyl-dihydroxyacetonephosphatesynthase by amyloid precursor protein processing and is affected in Alzheimer's disease. J Neurochem 116, 916-925.
- [25] Wood PL, Barnette BL, Kaye JA, Quinn JF, Woltjer RL (2015) Non-targeted lipidomics of CSF and frontal cortex grey and white matter in control, mild cognitive impairment, and Alzheimer's disease subjects. *Acta Neuropsychiatr* 27, 270-278.
- [26] Yamashita S, Kiko T, Fujiwara H, Hashimoto M, Nakagawa K, Kinoshita M, Furukawa K, Arai H, Miyazawa T (2016) Alterations in the levels of amyloid-β, phospholipid hydroperoxide, and plasmalogen in the blood of patients with Alzheimer's disease: Possible interactions between amyloid-β and these lipids. J Alzheimers Dis 50, 527-537.
- [27] Goodenowe DB, Cook LL, Liu J, Lu Y, Jayasinghe DA, Ahiahonu PWK, Heath D, Yamazaki Y, Flax J, Krenitsky KF, Sparks DL, Lerner A, Friedland RP, Kudo T, Kamino K, Morihara T, Takeda M, Wood PL (2007) Peripheral ethanolamine plasmalogen deficiency: A logical causative factor in Alzheimer's disease and dementia. *J Lipid Res* 48, 2485-2498.
- [28] Han X (2005) Lipid alterations in the earliest clinically recognizable stage of Alzheimer's disease: Implication of the role of lipids in the pathogenesis of Alzheimer's disease. *Curr Alzheimer Res* 2, 65-77.
- [29] Han X, Holtzman DM, McKeel Jr DW (2001) Plasmalogen deficiency in early Alzheimer's disease subjects and in animal models: Molecular characterization using electrospray ionization mass spectrometry. *J Neurochem* **77**, 1168-1180.
- [30] Wood PL, Mankidy R, Ritchie S, Heath D, Wood JA, Flax J, Goodenowe DB (2010) Circulating plasmalogen levels and Alzheimer Disease Assessment Scale-Cognitive scores in Alzheimer patients. J Psychiatry Neurosci **35**, 59-62.
- [31] Lim WLF, Huynh K, Chatterjee P, Martins I, Jayawardana KS, Giles C, Mellett NA, Laws SM, Bush AI, Rowe CC, Villemagne VL, Ames D, Drew BG, Masters CL, Meikle PJ, Martins RN (2020) Relationships between plasma lipids species, gender, risk factors, and Alzheimer's disease. J Alzheimers Dis 76, 303-315.
- [32] Toledo JB, Arnold M, Kastenmüller G, Chang R, Baillie RA, Han X, Thambisetty M, Tenenbaum JD, Suhre K, Thompson JW, John-Williams LS, MahmoudianDehkordi S, Rotroff DM, Jack JR, Motsinger-Reif A, Risacher SL, Blach C, Lucas JE, Massaro T, Louie G, Zhu H, Dallmann G, Klavins K, Koal T, Kim S, Nho K, Shen L, Casanova R, Varma S, Legido-Quigley C, Moseley MA, Zhu K, Henrion MYR, van der Lee SJ, Harms AC, Demirkan A, Hankemeier T, van Duijn CM, Trojanowski JQ, Shaw LM, Saykin AJ, Weiner MW, Doraiswamy PM, Kaddurah-Daouk R (2017) Metabolic network failures in Alzheimer's disease: A biochemical road map. *Alzheimers Dement* 13, 965-984.
- [33] Wykle RL (2004) Arachidonate remodeling and PAF synthesis in human neutrophils. In *Arachidonate Remodeling* and Inflammation, Birkhäuser, Basel, Basel, pp. 73-87.

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- [34] Prescott SM, Zimmerman GA, Stafforini DM, McIntyre TM 840 (2000) Platelet-activating factor and related lipid mediators. 841 Annu Rev Biochem 69, 419-445. 842
- [35] Han X, Holtzman DM, McKeel DW, Kelley J, Morris JC 843 844 (2002) Substantial sulfatide deficiency and ceramide elevation in very early Alzheimer's disease: Potential role in 845 disease pathogenesis. J Neurochem 82, 809-818. 846
- [36] Filippov V. Song MA, Zhang K, Vinters H V, Tung S, 847 848 Kirsch WM, Yang J, Duerksen-Hughes PJ (2012) Increased ceramide in brains with Alzheimer's and other neurodegen-849 erative diseases. J Alzheimers Dis 29, 537-547. 850
- [37] Han X, Rozen S, Boyle SH, Hellegers C, Cheng H, Burke 851 JR, Welsh-Bohmer KA, Doraiswamy PM, Kaddurah-Daouk 852 R (2011) Metabolomics in early Alzheimer's disease: Iden-853 tification of altered plasma sphingolipidome using shotgun lipidomics. PLoS One 6, e21643. 855

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- [38] He X, Huang Y, Li B, Gong CX, Schuchman EH (2010) Deregulation of sphingolipid metabolism in Alzheimer's disease. Neurobiol Aging 31, 398-408.
- Katsel P, Li C, Haroutunian V (2007) Gene expression [39] alterations in the sphingolipid metabolism pathways during progression of dementia and Alzheimer's disease: A shift toward ceramide accumulation at the earliest recognizable stages of Alzheimer's disease? Neurochem Res 32, 845-856.
- [40] Jana A, Hogan EL, Pahan K (2009) Ceramide and neurodegeneration: Susceptibility of neurons and oligodendrocytes to cell damage and death. J Neurol Sci 278, 5-15.
- [41] Trushina E, Mielke MM (2014) Recent advances in the application of metabolomics to Alzheimer's disease. Biochim Biophys Acta 1842, 1232-1239.
- Mielke MM, Bandaru VVR, Haughey NJ, Rabins P V., [42] 870 871 Lyketsos CG, Carlson MC (2010) Serum sphingomyelins 872 and ceramides are early predictors of memory impairment. Neurobiol Aging 31, 17-24. 873
- [43] Mapstone M, Cheema AK, Fiandaca MS, Zhong X, Mhyre 874 TR, Macarthur LH, Hall WJ, Fisher SG, Peterson DR, Haley 875 JM, Nazar MD, Rich SA, Berlau DJ, Peltz CB, Tan MT, 876 Kawas CH, Federoff HJ (2014) Plasma phospholipids iden-877 878 tify antecedent memory impairment in older adults. Nat Med 20, 415-418. 879
- [44] Casanova R, Varma S, Simpson B, Kim M, An Y, Saldana 880 S, Riveros C, Moscato P, Griswold M, Sonntag D, Wahrheit 881 J, Klavins K, Jonsson P V., Eiriksdottir G, Aspelund T, 882 Launer LJ, Gudnason V, Legido Quigley C, Thambisetty M 883 884 (2016) Blood metabolite markers of preclinical Alzheimer's disease in two longitudinally followed cohorts of older indi-885 viduals. Alzheimers Dement 12, 815-822. 886
- [45] Ma YH, Shen XN, Xu W, Huang YY, Li HQ, Tan L, Tan 887 CC, Dong Q, Tan L, Yu JT (2020) A panel of blood lipids 888 associated with cognitive performance, brain atrophy, and 889 Alzheimer's diagnosis: A longitudinal study of elders with-890 out dementia. Alzheimers Dement (Amst) 12, e12041. 891

5

- Li D, Misialek JR, Boerwinkle E, Gottesman RF, Shar-[46] rett AR, Moslev TH, Coresh J, Wruck LM, Knopman DS, Alonso A (2017) Prospective associations of plasma phospholipids and mild cognitive impairment/dementia among African Americans in the ARIC Neurocognitive Study. Alzheimers Dement (Amst) 6, 1-10.
- Varma VR, Oommen AM, Varma S, Casanova R, An [47] Y. Andrews RM, O'Brien R, Pletnikova O, Troncoso JC, Toledo J, Baillie R, Arnold M, Kastenmueller G, Nho K, Doraiswamy PM, Saykin AJ, Kaddurah-Daouk R, Legido-Quigley C, Thambisetty M (2018) Brain and blood metabolite signatures of pathology and progression in Alzheimer disease: A targeted metabolomics study. PLOS Med 15, e1002482.
- [48] Wood PL, Locke VA, Herling P, Passaro A, Vigna GB, Volpato S, Valacchi G, Cervellati C, Zuliani G (2016) Targeted lipidomics distinguishes patient subgroups in mild cognitive impairment (MCI) and late onset Alzheimer's disease (LOAD). BBA Clin 5, 25-28.
- [49] Barupal DK, Zhang Y, Fan S, Hazen SL, Tang WHW, Cajka T, Irvin MR, Arnett DK, Kind T, Kaddurah-Daouk R, Fiehn O (2019) The circulating lipidome is largely defined by sex descriptors in the GOLDN, GeneBank and the ADNI studies. bioRxiv 731448.
- [50] Arnold M, Nho K, Kueider-Paisley A, Massaro T, Huynh K, Brauner B, MahmoudianDehkordi S, Louie G, Moseley MA, Thompson JW, John-Williams LS, Tenenbaum JD, Blach C, Chang R, Brinton RD, Baillie R, Han X, Trojanowski JQ, Shaw LM, Martins R, Weiner MW, Trushina E, Toledo JB, Meikle PJ, Bennett DA, Krumsiek J, Doraiswamy PM, Saykin AJ, Kaddurah-Daouk R, Kastenmüller G (2020) Sex and APOE ɛ4 genotype modify the Alzheimer's disease serum metabolome. Nat Commun 11, 1148
- [51] Jiang Y, Zhu Z, Shi J, An Y, Zhang K, Wang Y, Li S, Jin L, Ye W, Cui M, Chen X (2019) Metabolomics in the development and progression of dementia: A systematic review. Front Neurosci 13, 343.
- [52] Dormann CF, Elith J, Bacher S, Buchmann C, Carl G, Carré G, Marquéz JRG, Gruber B, Lafourcade B, Leitão PJ, Münkemüller T, McClean C, Osborne PE, Reineking B, Schröder B, Skidmore AK, Zurell D, Lautenbach S (2013) Collinearity: A review of methods to deal with it and a simulation study evaluating their performance. Ecography (Cop) 36, 27-46.

935